

# Application Note

# GC–MS Air Analyser for the Determination of very low Concentrations of VOCs in Indoor and Ambient Air, Approaching method TO-17

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## **Key Words**

GC–MS, VOC, Air Quality , Air Analysis, Breath Analysis, Volatiles in Air, Ambient Air, Indoor Air, Airborne compounds, Volatile Organic Compounds.

### Abstract

An instrument has been developed for the automated, *in situ*, determination of airborne volatile organic compounds (VOCs) using programmed temperature vaporisation injection from a sorbent tube trap. A sorbent tube placed as an injection port liner can be repeatedly used to collect samples of air, with the trapped analytes being subsequently desorbed onto a capillary gas chromatography (GC) column without the use of intermediate cryogenic refocusing. The system does not need any liquid nitrogen or CO<sub>2</sub>, there is no need for adsorption/desorption tubes.



Graphical impression of the OPTIC – Air Analyser.



#### Introduction

Measurements of atmospheric concentrations of volatile organic compounds are currently required in both urban and rural environments, where their presence can be directly detrimental to health, and their ability to form photochemical oxidants in the presence of NOx and the sunlight is important. People breathe approximately 20 000 liters of air a day, so this concern is significant. It has also been demonstrated that some hydrocarbons may contribute to global warming and stratospheric ozone depletion and may also accumulate as persistent organic pollutants in some environments. Urban concentrations of VOCs are influenced directly by the emissions from major anthropogenic sources such as vehicle emissions, solvent and petrochemical evaporative losses, gas leaks, and many other forms of emission.

Many methods for analysing airborne VOCs have been proposed, typically using sorbent traps or evacuated canisters. Carbon molecular sieves of differing mesh sizes are particularly effective in trapping VOCs in air and are frequently incorporated into packed sorbent tubes. The analysis is usually then performed by thermally desorbing the collected sample in a stream of helium over a period of several minutes onto a cryogenically cooled intermediate trap. The sample is refocused at the intermediate trap prior to subsequent flash heating, thus introducing the collected analytes as a narrow band onto the analytical column. A method based on a thermally desorbed adsorption trap and cryogenic refocusing with liquid nitrogen (LN<sub>2</sub>) or carbon dioxide (CO<sub>2</sub>) uses typically 50 liters of coolant per day. Whilst methods involving the cryogenic refocusing of sample produce effective analytical separations, the equipment required is complex and has high operation costs. Also, such a device requires regular user attention to replenish cryogen supplies.



Mobile lab for *in situ* VOC analysis.

Placing a sorbent tube as an injection port liner inside a programmed temperature vaporisation (PTV) injector allows rapid desorption of analytes directly into an analytical column. Using this method the traditional intermediate refocusing between desorption and separation is no longer required.

In this application note we describe the use of an online sorbent tube sample collection, with programmed temperature vaporisation (PTV) injector for the *in situ* analysis of ambient air samples. The automation of sample collection and the elimination of the intermediate refocusing step, reduces much of the necessary on-site maintenance required for continuous monitoring, and allows *in situ* analysis in locations where previously this was not possible.



Using a multi positioning valve, the system can select between standards from, *e.g.*, a Tedlar<sup>®</sup> bag or taking sample directly from the outside via, e.g., a probe on the roof of a mobile lab. Via a smart flow system, a sample is pumped through the injector port liner containing a suitable trapping material in reverse flow from bottom to top trapping the analytes, while the carrier gas is rerouted to the GC capillary column bypassing the injector port. Sampling is done during a fixed time with a constant flow controlled with a mass flow controller, without the use of additional coolant as LN<sub>2</sub> or CO<sub>2</sub>.

Once the sampling period is finished, the carrier gas flow via the injector port is reestablished. Following this, the injection port is heated to desorb the analytes from the injection port liner for transfer to the GC capillary column. In parallel with heating the injection port liner, the GC–MS analysis is started.

#### Experimental

The instruments used in this application are the OPTIC Air Analyser (GL Sciences, The Netherlands), containing an OPTIC AirLiner (GL Sciences), and the QP2020 NX GC–MS (Shimadzu, Japan). Detailed analytical parameters are described in table 1. For the chromatography an Agilent PoraBOND Q capillary column ( $25 \text{ m x } 0.25 \text{ mm x } 3 \mu m$ ) was used. This column was chosen because it focuses and separates the volatiles efficiently, in addition with an adequate temperature range to elute the highest boiling compounds.



Table 1: Analytical Parameters

OPTIC Air Analyser Parameters		GC–MS Parameters		
Sampling Temp	25°C	Initial Oven Temp	30°C	
Sampling Flow	75 ml/min	Initial time	2.50 min	
Sampling Time	3.00 min	Rate 1	14°C/min	
Desorption Temp	270°C	Final Temp	270°C	
Desorption Temp Hold	270 sec	Final Time	5.36 min	
Transfer Time	220 sec	Total runtime	25 min	
Column Carrier gas	Helium			
Column Flow	1.6 ml/min	MS Mode	Scan / SIM	

#### Results

A home-made mixture of 63 compounds (comparable to the TO-17 mixture) is used for evaluation of the Air Analyser. Via a smart valving system, sample is loaded from bottom-totop on the OPTIC Air Liner, being located inside the injection port at a temperature of 25°C The injection port is cooled with compressed air. Sampling is done during three minutes at 75 ml/min (total sample volume = 225 ml). After sampling, the carrier gas is re-routed through the injection port from top-to-bottom. Following this, the injection port is heated to 270°C, desorbing the trapped compounds to the capillary column using splitless transfer. The resulting chromatogram starts with *n*-Propane and ends with Naphthalene. Since the sample trap is in the injection port, no additional cryogenic focusing is required at the head of the capillary column.



Figure 1: TIC chromatogram (scan) of a 63 compound mixture at 25  $\mu$ g/m<sup>3</sup>.



The precision, linearity and instrument detection limit (IDL) is excellent for all compounds (see table 2). The precision was investigated by 10 consecutive runs of a standard with an average concentration of 0.25  $\mu$ g/m<sup>3</sup>. The linearity was determined by running standards with average concentrations in the range of 0.25 – 25  $\mu$ g/m<sup>3</sup>. For IDL determination, eight consecutive runs of a standard just above the limit are performed. Following the IDL is calculated using the student's t-test with a confidence level of 98% (t=2.896).Because of the relatively small amount of packing material in the OPTIC AirLiner, it was tested if whether the sampling conditions ( 3 min, 75 ml/min, 25°C) are close to the breakthrough volume. For this, sampling was done of the 2.5- $\mu$ g/m<sup>3</sup> standard for 1 – 10 minutes (75 – 750 ml). In figure 2, peak areas of some important compounds are given as a function of the sampling volume up to 750 ml.





#### Additional options to the system

- In case of high humidity samples, a gas dryer can get implemented in the system. Moisture will have an influence on the capacity of the trapping material in the injection port liner.
- In case of off-line loading of liners, the system can become upgraded with a manual or automatic liner exchange tool. This makes to system extremely flexible in its applicability.

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#### Conclusions

- An analyser for *in situ* Analysis of VOCs in Urban Air has been demonstrated.
- Air samples are automatically enriched on a sorbent tube in a GC injection port and subsequently analysed by GC–MS.
- No need for LN<sub>2</sub> or CO<sub>2</sub> as trapping coolant.
- Instrument Detection Limit (IDL) is for the majority of the compounds in the range of 10 – 100 ng/m<sup>3</sup>.

Name	t <sub>R</sub> (min)	Quant Masses	Corr. Coeff.	IDL (ng/m <sup>3</sup> )	%RSD*
Propane	5.97	43	0.996	72.9	7.8
Ethene, chloro-	7.22	62	0.997	36.0	13.6
Ethanol	8.13	45	0.862	27.5	3.7
Isobutane	8.53	43	0.992	39.8	4.2
1,3-butadiene	8.58	54	0.945	42.0	4.8
1-Butyne	8.77	54	1.000	87.4	9.9
Butane	8.99	43	0.841	67.7	7.1
Methylene chloride	9.99	84	0.999	90.8	12.1
Propenenitrile	10.08	53	0.992	12.8	2.9
Isopropyl Alcohol	10.14	45	0.628	16.8	7.0
Carbon disulfide	10.16	78	0.990	168.6	10.1
Dimethyl sulfide	10.31	62	0.995	98.7	7.2
Ethene, 1,1-dichloro-	10.41	96	1.000	20.7	3.0
1-Propanol	10.85	59	0.998	71.5	9.8
trans-1,2-Dichloroethylene	11.06	96	1.000	30.5	2.1
Diethyl ether	11.08	74	1.000	22.5	3.5
Isoprene	11.20	67	0.999	37.0	9.6
Pentane	11.44	72	0.998	25.2	6.5
cis-1,2-Dichloroethylene	11.74	96	1.000	16.1	2.2
Ethane, 1,1-dichloro-	11.82	63	0.999	61.2	9.2
Butanone	12.26	72	0.989	57.4	7.9
Trichloromethane	12.26	83	0.999	322.8	38.5
Tetrahydrofuran	12.32	71	0.999	192.1	23.8
2-Butanol	12.50	59	0.999	33.5	4.6
Ethane, 1,2-dichloro-	12.79	62	0.999	28.7	4.1
Methyl t-butyl ether (MTBE)	12.84	73	0.994	87.3	13.0
1-Butanol	13.17	31	0.997	54.5	7.4
Benzene	13.45	78	0.994	16.1	3.2
Ethane, 1,1,1-trichloro-	13.50	97	1.000	20.4	2.7

#### Table 2: Analytical results.

#### Table 2: (Continued)

Name	t <sub>R</sub> (min)	Quant Masses	Corr. Coeff.	IDL (ng/m3)	%RSD
Cyclopentane, methyl-	13.53	69	0.999	37.6	8.9
n-Hexane	13.54	86	1.000	65.2	16.8
Carbon Tetrachloride	13.61	119	0.999	27.9	3.1
Trichloroethylene	13.63	95	0.995	18.7	2.3
Cyclohexane	13.98	84	0.999	228.3	51.8
Propane, 1,1-dichloro-	14.03	77	0.997	94.1	14.7
Ethyl tert-butyl ether (ETBE)	14.13	87	0.998	18.9	2.8
Propane, 1,2-dichloro-	14.33	63	1.000	56.5	8.6
Ethane, 1,1,2-trichloro-	15.10	97	1.000	46.8	5.8
Propane, 1,3-dichloro-	15.25	76	0.999	46.5	6.9
Heptane	15.42	43	0.998	46.8	12.1
Toluene	15.45	65	0.938	56.7	11.6
Tetrachloroethylene	15.52	166	1.000	55.3	6.0
Benzene, chloro-	16.34	112	0.999	12.0	1.9
Bromoform	16.85	173	0.994	209.9	12.8
4-Vinyl-1-cyclohexene	16.99	79	0.999	52.9	11.2
Octane	17.05	57	0.979	43.0	10.8
Ethylbenzene	17.07	106	0.968	40.1	8.1
m,p-Xylene	17.11	106	0.776	140.7	14.4
o-Xylene	17.34	106	0.990	77.3	15.5
Benzene, 1,3-dichloro-	18.35	146	0.998	32.3	4.4
Nonane	18.40	57	0.987	101.2	24.9
Benzene, 1,3,5-trimethyl	18.43	92	0.999	51.3	10.5
Benzene, 1,3,5-trimethyl	18.43	105	0.991	54.0	11.1
Benzene, 1,2-dichloro-	18.45	146	0.998	18.5	2.5
Benzene, 1,4-dichloro-	18.65	146	0.999	49.2	8.7
Benzene, 1,2,4-trimethyl-	18.69	120	0.977	37.0	7.5
Indane	18.93	118	0.996	68.2	12.1
Decane	19.87	142	0.989	25.4	6.2
Decaline	20.54	96	0.999	76.7	15.1
Benzyl methyl ketone	20.58	134	0.705	39.8	4.4
Benzene, 1,2,4-trichloro-	20.77	180	0.994	24.2	2.9
Naphthalene	21.64	128	0.962	17.5	3.1
Undecane	22.03	57	0.815	11.0	2.6

\* Calculation based on measurements close to the IDL level.

$$IDL = \frac{StDev}{Average \ peak \ area} * concentration * t$$